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Summarising and validating test accuracy results across multiple studies for use in clinical practice

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Following a meta-analysis of test accuracy studies, the translation of summary results into clinical practice is potentially problematic. The sensitivity, specificity and positive (PPV) and negative (NPV) predictive values of a test may differ substantially from the average meta-analysis findings, because of heterogeneity. Clinicians thus need more guidance: given the meta-analysis, is a test likely to be useful in new populations, and if so, how should test results inform the probability of existing disease (for a diagnostic test) or future adverse outcome (for a prognostic test)? We propose ways to address this. Firstly, following a meta-analysis, we suggest deriving prediction intervals and probability statements about the potential accuracy of a test in a new population. Secondly, we suggest strategies on how clinicians should derive post-test probabilities (PPV and NPV) in a new population based on existing meta-analysis results and propose a cross-validation approach for examining and comparing their calibration performance. Application is made to two clinical examples. In the first example, the joint probability that *both* sensitivity and specificity will be >80% in a new population is just 0.19, because of a low sensitivity. However, the summary PPV of 0.97 is high and calibrates well in new populations, with a probability of 0.78 that the true PPV will be at least 0.95. In the second example, post-test probabilities calibrate better when tailored to the prevalence in the new population, with cross-validation revealing a probability of 0.97 that the observed NPV will be within 10% of the predicted NPV. © 2015 The Authors. *Statistics in Medicine* Published by John Wiley & Sons Ltd.

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1. Introduction

Test accuracy studies aim to evaluate the performance of a candidate medical test for either diagnosing the presence of a particular clinical condition ('diagnostic test') or identifying those likely to experience a particular outcome in the future ('prognostic test'). Such tests include measurable variables such as biomarkers, blood pressure and temperature, or may reflect a clinician's judgement after a physical examination or imaging result. When multiple studies evaluate the performance of a potential diagnostic or prognostic test, meta-analysis methods can synthesise the study results to help establish if and how

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the test can be used in practice [1–4]. Current meta-analyses of test accuracy studies focus on producing pooled estimates that summarise *average* test performance across the multiple studies [2–6]. This typically leads to a single pooled estimate for each of sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) and sometimes a summary ROC curve. For example, in patients undergoing a thyroidectomy, Noordzij *et al.* [7] examine the accuracy of the % change in parathyroid (PTH) levels (from pre-surgery to post-surgery) for identifying those at high risk of becoming hypocalcemic within 48 h. They synthesise data from six studies and summarise that a 70% or greater reduction in PTH has a sensitivity of 0.93, a specificity of 0.88, a PPV of 0.74 and an NPV of 0.97. However, when data are available from multiple studies, a major concern is heterogeneity in test accuracy, which refers to genuine differences in the accuracy of a test from study to study. Heterogeneity in test accuracy is common in meta-analysis and is caused by study differences in, for example, clinical settings (such as hospitals), patient selection (such as symptom severity and previous testing), choice of health-care practitioners implementing/reading the test, the included patient populations (e.g. with a different underlying prevalence of disease) and the test methodology employed (e.g. choice of threshold, method of measurement and the reference standard).

Where heterogeneity exists, this is usually accounted for by using random-effects models that allow test accuracy to vary across studies [2, 4]. However, typically, the aim of meta-analysis is as before: to estimate the *average* test performance across all the populations included. Although this is a worthwhile research question, the translation of such *average* meta-analysis results into clinical practice is potentially problematic. For example, a diagnostic test may have high sensitivity and specificity values on average, but the causes of heterogeneity may lead to substantially lower values in some particular populations, leading to poor performance. Similarly, average post-test probabilities from a prognostic test may be inaccurate, for example, if the outcome prevalence in a particular population is markedly different than the average outcome prevalence across all populations. For example, Leeftang *et al.* [5] said ‘when a review contains studies with a broad range of prevalence, there will be difficulties in working out how the average predictive values (estimated for the average included study with an average prevalence) can be applied in clinical practice.’

To address this concern, this article suggests ways to improve the translation of meta-analysis results for clinical practice, by going beyond presenting just average test accuracy results. There are two main objectives. Firstly, we emphasise the importance of deriving and reporting prediction intervals for the potential accuracy of a test in a new population [8, 9]. Secondly, we propose a cross-validation approach for examining how to derive PPV and NPV in new populations based on existing meta-analysis results. Our viewpoint is similar to Willis and Hyde [10], who propose that meta-analysis results need to be tailored before use in a particular population, and Leeftang *et al.* [5], who propose predicting PPV and NPV in new populations. Our cross-validation framework provides a way to examine the calibration of approaches (or statistical equations) that tailor (or predict) results for new populations.

The remainder of the paper is as follows. In Section 2, we introduce two motivating datasets that are used throughout the article. Section 3 introduces the derivation and interpretation of prediction intervals (regions) following meta-analysis, in terms of the discrimination ability of a test (sensitivity, specificity and *c* statistic). Section 4 suggests approaches to derive PPV and NPV in new populations and outlines the cross-validation framework for examining their calibration. Section 5 considers extension to comparing tests. Section 6 provides some discussion, and Section 7 concludes.

2. Motivating datasets

2.1. Accuracy of ear temperature for diagnosing fever in children

Craig *et al.* [11] systematically reviewed thermometry studies comparing temperatures taken at the ear and rectum in children, and of clinical interest is the accuracy of infrared ear thermometry for diagnosing fever [12]. Eleven studies (involving 2323 children) evaluated the accuracy of a ‘FirstTemp’ branded ear thermometer in relation to an electronic rectal thermometer. Rectal temperature was the reference measure, as it is a well-established method of measuring temperature in children, and it guides clinical decisions and the definition of fever. However, measuring temperature at the ear is clearly more acceptable than measuring temperature at the rectum, and so ear measurements would be preferable if their diagnostic accuracy is adequate. All studies defined patients with an ear temperature ≥ 38.0 °C to be test positive, and the gold standard definition of fever was a rectal temperature ≥ 38.0 °C (Table I), consistent with National Health Service guidelines for diagnosing fever in children at the time of these studies.

Table I. Summary of the 11 temperature studies identified for meta-analysis; each study used a threshold of 38 °C to define fever, an electronic rectal thermometer and a FirstTemp ear thermometer.

| First author | r_{11i} | n_{1i} | Sensitivity | r_{00i} | n_{0i} | Specificity | Observed prevalence |
|--------------|-----------|----------|-------------|-----------|----------|-------------|---------------------|
| Brennan | 150 | 203 | 0.74 | 155 | 167 | 0.93 | 0.55 |
| Davis | 9 | 18 | 0.50 | 46 | 48 | 0.96 | 0.27 |
| Green | 8 | 9 | 0.89 | 12 | 12 | 1.00 | 0.43 |
| Greenes | 53 | 109 | 0.49 | 193 | 195 | 0.99 | 0.36 |
| Hoffman | 30 | 42 | 0.71 | 56 | 58 | 0.97 | 0.42 |
| Hooker | 10 | 15 | 0.67 | 24 | 24 | 1.00 | 0.38 |
| Lanham | 53 | 103 | 0.51 | 74 | 75 | 0.99 | 0.58 |
| Muma | 48 | 87 | 0.55 | 136 | 136 | 1.00 | 0.39 |
| Nypaver | 282 | 425 | 0.66 | 445 | 453 | 0.98 | 0.48 |
| Rhoads | 7 | 27 | 0.26 | 38 | 38 | 1.00 | 0.42 |
| Stewart | 57 | 59 | 0.97 | 20 | 20 | 1.00 | 0.75 |

r_{11i} , number of true positives; n_{1i} , number with fever.

r_{00i} , number of true negatives; n_{0i} , number without fever.

Table II. Summary of five cohort studies evaluating the accuracy of a >65% change in PTH (measured pre-operation to 0–20 min post-thyroidectomy) for identifying hypocalcaemia.

| First author | r_{11i} | n_{1i} | Sensitivity | r_{00i} | n_{0i} | Specificity | Observed proportion with outcome |
|--------------|-----------|----------|-------------|-----------|----------|-------------|----------------------------------|
| Lo | 11 | 11 | 1.0 | 56 | 89 | 0.63 | 0.11 |
| Lombardi | 13 | 16 | 0.81 | 31 | 35 | 0.89 | 0.31 |
| McLeod | 9 | 13 | 0.69 | 33 | 43 | 0.77 | 0.23 |
| Warren, 2002 | 3 | 4 | 0.75 | 10 | 12 | 0.83 | 0.25 |
| Warren, 2004 | 2 | 3 | 0.67 | 20 | 23 | 0.87 | 0.12 |

r_{11i} , number of true positives; n_{1i} , number with hypocalcaemia.

r_{00i} , number of true negatives; n_{0i} , number without for identifying hypocalcaemia.

PTH, parathyroid.

Table III. Summary of five cohort studies evaluating the accuracy of a >65% change in PTH (measured pre-operation to 1–2 h post-thyroidectomy) for identifying hypocalcaemia.

| First author | r_{11i} | n_{1i} | Sensitivity | r_{00i} | n_{0i} | Specificity | Observed proportion with outcome |
|--------------|-----------|----------|-------------|-----------|----------|-------------|----------------------------------|
| Lam | 12 | 12 | 1.0 | 24 | 26 | 0.92 | 0.32 |
| Lombardi | 15 | 16 | 0.94 | 29 | 35 | 0.83 | 0.31 |
| McLeod | 11 | 12 | 0.92 | 19 | 25 | 0.76 | 0.32 |
| Warren, 2002 | 1 | 2 | 0.50 | 6 | 8 | 0.75 | 0.2 |
| Warren, 2004 | 3 | 3 | 1.0 | 17 | 23 | 0.74 | 0.12 |

r_{11i} , number of true positives; n_{1i} , number with hypocalcaemia.

r_{00i} , number of true negatives; n_{0i} , number without hypocalcaemia.

PTH, parathyroid.

The studies included children already in hospital or attending accident and emergency [11], and so the observed prevalence of fever was high, around 50%. The summary results for each study are shown in Table I in terms of the totals with and without fever, estimated sensitivity and specificity and the number of true positives and negatives. Meta-analysis is required to summarise test accuracy across these studies, to appropriately quantify any between-study heterogeneity in test accuracy and – most important to this article – to identify the potential performance of the test in new populations.

2.2. Accuracy of % change in PTH for predicting risk of hypocalcaemia

As mentioned, Noordzij *et al.* [7] examined the accuracy of the % change in PTH for identifying patients at high risk of becoming hypocalcemic within 48 h after a thyroidectomy. The % change from baseline (rather than absolute change) was used to minimise the impact of different methods of measuring PTH across studies. We focus here on the accuracy of PTH when using a threshold of 65%, above which a patient is classed as test positive. Table II summarises the test accuracy results for five cohort studies for % change in PTH when measured from baseline (before surgery) to 0–20 min post-surgery. Similarly, Table III summarises the five cohort studies evaluating % change in PTH when measured from baseline to 1–2 h post-surgery. Meta-analysis results are important here to inform whether PTH can suitably identify those at high risk of hypocalcaemia (to ensure they remain in hospital) and those at low risk of hypocalcaemia (who may therefore benefit from early hospital discharge). If PTH appears useful, understanding how best to translate the meta-analysis results into clinical practice is then crucial.

3. Examining and summarising the discrimination of a test using meta-analysis

Discrimination statistics, such as sensitivity and specificity, quantify how a test distinguishes between patients with and without the disease (or outcome) of interest and are thus conditional on knowing true disease status. In this section, we show how to use meta-analysis results for predicting a test's discrimination performance in a new population. For simplicity, we refer only to *diagnostic* tests, but the same principles apply for prognostic test studies where outcome status is known for all patients by a given time.

3.1. Sensitivity and specificity

3.1.1. Bivariate meta-analysis model. Suppose there are k test accuracy studies (indexed by i , where $i = 1$ to k) including n_{1i} and n_{0i} patients with and without disease, respectively. The test classifies each patient as either positive or negative, with the aim that those positive are truly diseased and those negative are truly non-diseased. Summarising test results over all patients in each study produces aggregate data in the form of r_{11i} (true positives), the number of patients in study i with a positive test result who truly have the disease, and r_{00i} (true negatives), the number of patients in study i with a negative test result who truly do not have the disease (true negatives). The observed sensitivity in each study is r_{11i}/n_{1i} , and the observed specificity is r_{00i}/n_{0i} . In this situation, a bivariate random-effects meta-analysis can jointly synthesise sensitivity and specificity across studies [2, 3], with r_{11i} and r_{00i} modelled directly using the binomial distribution [4, 13] and allowing for potential between-study heterogeneity in logit-sensitivity and logit-specificity, and their potential between-study correlation due to explicit and implicit differences in threshold value across studies [2].

$$\begin{aligned} r_{11i} &\sim \text{Binomial}(n_{1i}, p_{1i}) \\ r_{00i} &\sim \text{Binomial}(n_{0i}, p_{0i}) \\ \begin{pmatrix} \text{logit}(p_{1i}) \\ \text{logit}(p_{0i}) \end{pmatrix} &\sim N \left[\begin{pmatrix} \beta_1 \\ \beta_0 \end{pmatrix}, \mathbf{\Omega} \right], \mathbf{\Omega} = \begin{pmatrix} \tau_1^2 & \tau_1 \tau_0 \rho \\ \tau_1 \tau_0 \rho & \tau_0^2 \end{pmatrix} \end{aligned} \quad (1)$$

Model (1) specifies an underlying logit-sensitivity and logit-specificity in each study, by $\text{logit}(p_{1i})$ and $\text{logit}(p_{0i})$, respectively, which are assumed normally distributed about a mean logit-sensitivity of β_1 and a mean logit-specificity of β_0 , with between-study variances τ_1^2 and τ_0^2 , respectively, and between-study correlation ρ . Estimation of model (1) is possible in both frequentist and Bayesian frameworks, using maximum likelihood estimation via Gaussian quadrature [4, 14–16] or MCMC estimation using Gibbs sampling, respectively [17–20]. For brevity, we present full estimation details in the Supporting Information (S1 and S2).

3.1.2. Summarising discrimination for clinical practice. When reporting results from model (1), researchers typically focus on the summary estimates of sensitivity and specificity, obtained by $\exp(\hat{\beta}_1)/[1 + \exp(\hat{\beta}_1)]$ and $\exp(\hat{\beta}_0)/[1 + \exp(\hat{\beta}_0)]$, respectively, and their CIs (or joint confidence region) [2]. However, because of heterogeneity, these summary results may be misleading as they do not necessarily reflect the test's sensitivity and specificity when applied to particular populations. We therefore suggest that researchers should also calculate and report

- (1) a prediction region for the true sensitivity and specificity in a new population [2], such as a 95% prediction region; and
- (2) the probability that the true sensitivity and specificity will *both* be above some clinically acceptable values in the new population.

These measures reveal the potential impact of heterogeneity on discrimination performance in new populations and thereby help indicate whether a test is fit for purpose. For example, if both sensitivity and specificity need to be at least 90% for the test to be considered useful, there should be a large probability that sensitivity and specificity will both meet this criteria in a new population. To make these inferences following estimation of model (1), the joint predictive distribution for a new pair of sensitivity and specificity needs to be derived. If β_1 , β_0 , τ_1^2 , τ_0^2 and ρ were known, then this distribution on the logit-scale would be

$$\begin{pmatrix} \text{logit}(p_{1i})_{NEW} \\ \text{logit}(p_{0i})_{NEW} \end{pmatrix} \sim N \left[\begin{pmatrix} \beta_1 \\ \beta_0 \end{pmatrix}, \mathbf{\Omega} \right], \mathbf{\Omega} = \begin{pmatrix} \tau_1^2 & \tau_1 \tau_0 \rho \\ \tau_1 \tau_0 \rho & \tau_0^2 \end{pmatrix} \quad (2)$$

However, these parameters are not known, and so the predictions should also account for their uncertainty. This is most natural in a Bayesian framework [8], where the uncertainty in all parameters estimated in model (1) is propagated when deriving predictions for new sensitivities and specificities. For our Bayesian application of model (1) in this paper, vague prior distributions of $N(0, 100\,000)$ were specified for β_1 and β_0 , and a Wishart prior distribution for $\mathbf{\Omega}^{-1}$ (Supporting Information S2). However, alternative priors for the components of $\mathbf{\Omega}$ are also possible [21, 22]. During the Bayesian estimation, posterior inferences about $\text{logit}(p_{1i})_{NEW}$ and $\text{logit}(p_{0i})_{NEW}$ can be made by sampling from their joint posterior (predictive) distribution to calculate 95% intervals and regions, and joint probability statements.

In a frequentist framework, a bivariate t -distribution with $k - 2$ degrees of freedom might be used to approximate the joint predictive distribution, which extends the frequentist use of a univariate t -distribution to derive a prediction interval for a single measure [8, 9]. For example, a prediction interval for $\text{logit}(p_{1i})_{NEW}$ is

$$\hat{\beta}_1 - t_{k-2} \sqrt{\hat{\tau}_1^2 + \text{Var}(\hat{\beta}_1)}, \hat{\beta}_1 + t_{k-2} \sqrt{\hat{\tau}_1^2 + \text{Var}(\hat{\beta}_1)} \quad (3)$$

where $\text{Var}(\hat{\beta}_1)$ is the variance of $\hat{\beta}_1$ and t_{k-2} is the $100(1 - \alpha/2)$ percentile of the t -distribution with $k - 2$ degrees of freedom, with α usually chosen as 0.05 to give a 95% prediction interval. A t -distribution, rather than a normal distribution, is used to help account for the uncertainty in $\hat{\tau}_1^2$.

3.2. c Statistic

Tests measured on a continuous scale (such as PTH) allow sensitivity and specificity to be estimated at each of multiple thresholds to define test positive and test negative and thus produce an ROC curve. In this situation, a measure of the test's discrimination across all thresholds is the area under the ROC curve, otherwise known as the c statistic [23]. This gives the probability that, if one diseased person and one non-diseased person were chosen at random, the continuous test value would be larger for the diseased person. Given multiple studies, Van Klaveren *et al.* [24] propose that a random-effects meta-analysis of the c statistic estimates is useful:

$$\begin{aligned} c_i &\sim N(\theta_i, s_i^2) \\ \theta_i &\sim N(\mu_c, \tau_c^2) \end{aligned} \quad (4)$$

Here, c_i is the c estimate in study i assumed to have an (approximate) normal sampling distribution with its variance, s_i^2 , assumed known and its mean, θ_i , the true c statistic in study i , which itself is assumed drawn from a normal distribution with summary mean μ_c and between-study variance τ_c^2 . When the raw test data are available from each study, the c_i can easily be obtained using standard statistical packages, and most typically, bootstrapping can be used to obtain the s_i^2 required. Model (4) can be estimated in a frequentist framework, for example, using methods of moments or restricted maximum likelihood, and the 95% prediction interval is then derived using

$$\hat{\mu}_c - t_{k-2} \sqrt{\hat{\tau}_c^2 + \text{Var}(\hat{\mu}_c)}, \hat{\mu}_c + t_{k-2} \sqrt{\hat{\tau}_c^2 + \text{Var}(\hat{\mu}_c)} \quad (5)$$

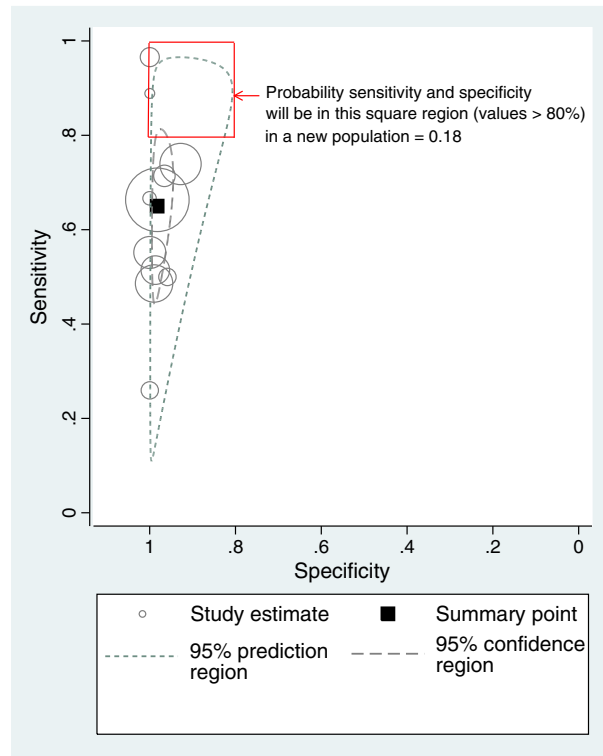


Figure 1. Confidence and prediction regions following application of model (1) to the temperature data.

where $\text{Var}(\hat{\mu}_c)$ is the variance of $\hat{\mu}_c$ and other terms follow as outlined in Equation (3). In a Bayesian framework, (vague) prior distributions are required, such as $\mu_c \sim U(0.5, 1)$ and $\tau_c \sim U(0, 0.25)$, and then probability statements about the potential c statistic in a new population can be obtained from the posterior distribution for θ_{NEW} , such as the probability that the c statistic will be above 0.9.

3.3. Application

3.3.1. Ear temperature for diagnosis of fever. Model (1) was fitted to the 11 studies evaluating an ear temperature $\geq 38.0^\circ\text{C}$ for identifying fever in children (Table I). The frequentist results are displayed in Figure 1 and are very similar to the results from a Bayesian approach, which we now discuss. The summary sensitivity is 0.65 (95% CrI: 0.51 to 0.77) and the summary specificity is 0.98 (95% CrI: 0.96 to 0.99). The summary specificity is high, so those without fever are usually correctly classed as negative ($< 38.0^\circ\text{C}$). However, the summary sensitivity is low, indicating that many children with fever are not classed as positive ($\geq 38.0^\circ\text{C}$). Furthermore, despite the same ear thermometer manufacturer (first-temp), a consistent threshold (38.0°C) and a common reference standard (rectal temperature), there is considerable between-study heterogeneity in both sensitivity and specificity. This is illustrated by the wide prediction region within Figure 1. A 95% prediction interval for sensitivity in a new population is calculated as 0.21 to 0.94 and for specificity is calculated as 0.89 to 1. Direct probabilistic statements can also be made from the (joint) predictive distribution for a new sensitivity and specificity (Supporting Information S3). For example, in a new population, the probability that sensitivity will be over 80% is 0.19, and the probability that specificity will be over 80% is 0.99. Further, the joint probability that in a new population *both* sensitivity and specificity will be $> 80\%$ is just 0.19 (Figure 1). Hence, if sensitivity and specificity above 80% are deemed the minimum acceptable, there is less than a one in five chance that ear temperature will be adequate within a new population. So there is little evidence here to suggest that ear temperature can replace rectal temperature as an acceptable technique for assessing fever.

3.3.2. PTH for predicting hypocalcaemia. Parathyroid is measured on a continuous scale, and as the raw data were available to us, we calculated the c statistic and its 95% CI and bootstrap standard error for each of the five studies. Model (4) was then estimated using methods of moments [25], and the results are shown in the Supporting Information (S4) for the test relating to the % change in PTH from baseline

to 0–20 min post-surgery. The summary c statistic is encouragingly high (0.92, 95% CI: 0.86 to 0.97). Furthermore, although there is heterogeneity in discrimination performance across studies ($I^2 = 28\%$), the 95% prediction interval derived from Equation (5) indicates that c in a new population will be between 0.78 and 1.00, and is thus also predominately high. Bayesian inferences were similar, but the width of intervals were sensitive to the prior distribution for the between-study variance. However, for all the priors considered, there was a probability of about 0.89 that c in a new population will be above 0.8 and a probability of 0.59 that it will be above 0.9. Therefore, there is large probability that PTH measured at 0–20 min will have very good discrimination when applied in new populations. The discrimination was similar when PTH was measured 1–2 h post-surgery.

For implementation of PTH in clinical practice, a single threshold would be chosen to define test positive and negative. Noordzij *et al.* [7] recommend a threshold of a 65% change in PTH from pre-surgery to 1–2 h post-surgery, for which Bayesian estimation of model (1) gives a summary sensitivity of 0.93 (95% CrI: 0.83 to 0.99) and a summary specificity of 0.81 (95% CrI 0.71 to 0.89). The probability that sensitivity will be greater than 0.8 in a new population is 0.96, and for specificity it is 0.59 (Supporting Information S3). The probability that both will be above 0.8 is 0.57. In other words, we expect that in nearly 6 out of 10 populations, both the sensitivity and specificity of the PTH test will be above 0.8. Thus, PTH measured at 1–2 h appears to have high discrimination for potential clinical use, and this is evaluated further in terms of post-test predictions in Section 4.3.

4. Examining and summarising the calibration of post-test probabilities derived following meta-analysis

In practice, clinicians are more interested in PPV and NPV, the post-test disease (outcome) probabilities conditional on knowing the test result. Following meta-analysis, an explicit strategy (or statistical equation) is therefore needed to tell clinicians how to derive PPV and NPV in their population. Further, any recommended strategy or equation should be formally evaluated in new data, a concept known as *external validation* in the prediction modelling field [26]. Thus, in this section, we consider (i) strategies for deriving PPV and NPV in new populations using existing meta-analysis results, and then (ii) an approach called internal–external cross-validation [27, 28], which allows the strategies to be examined and compared in terms of calibration of predicted and observed PPV and NPV.

4.1. Strategies to derive PPV and NPV in new populations

For simplicity, let us focus on prognostic test studies, as these are usually cohort studies for which absolute risk can be observed [29]. Let there be n_{1i} and n_{0i} patients that do and do not have an outcome by a particular time, respectively, and let r_{11i} be the number of patients in study i who had the outcome and a positive test result at baseline, and r_{00i} be the number of patients in study i who did not have an outcome and had a negative test result at baseline. We now suggest three possible strategies for deriving PPV and NPV in a new population (L), given k existing studies for meta-analysis.

4.1.1. Modelling option A: use summary values of sensitivity, specificity and prevalence. The simplest approach is to derive the new PPV and NPV by first obtaining meta-analysis summary estimates of sensitivity, specificity and prevalence, and then plugging them into the well-known Bayes theorem solutions for deriving PPV and NPV from sensitivity, specificity and prevalence:

$$\text{PPV}_{\text{new}_L} = \frac{\text{sensitivity}_{\text{MA}} \times \text{prevalence}_{\text{MA}}}{[\text{sensitivity}_{\text{MA}} \times \text{prevalence}_{\text{MA}}] + [(1 - \text{specificity}_{\text{MA}})(1 - \text{prevalence}_{\text{MA}})]} \quad (6)$$

$$\text{NPV}_{\text{new}_L} = \frac{\text{specificity}_{\text{MA}} \times (1 - \text{prevalence}_{\text{MA}})}{[(1 - \text{sensitivity}_{\text{MA}}) \times \text{prevalence}_{\text{MA}}] + [\text{specificity}_{\text{MA}} \times (1 - \text{prevalence}_{\text{MA}})]} \quad (7)$$

In these equations, ‘MA’ denotes the summary meta-analysis value, and ‘new’ denotes the predicted value in the new population, L . This approach mimics situations where nothing is known about test accuracy performance and prevalence in the new population, and so one simply takes the summary values from existing studies. The summary sensitivity and summary specificity can be obtained from fitting model

(1) to the available studies (which may include cohort and case-control studies), and the summary prevalence can be obtained from a meta-analysis of available cohort studies, for example, by using maximum likelihood to fit:

$$\begin{aligned} n_{1i} &\sim \text{Binomial}(N_i, \text{prevalence}_i) \\ \text{logit}(\text{prevalence}_i) &\sim N(\alpha, \tau_\alpha^2) \end{aligned} \quad (8)$$

where N_i is the total sample size in study i and the summary prevalence is obtained by $\exp(\hat{\alpha})/(1+\exp(\hat{\alpha}))$.

4.1.2. Modelling option B: use summary values of sensitivity and specificity, and known prevalence in the intended population. Even when there is no between-study heterogeneity in sensitivity and specificity, it is likely that there is heterogeneity in the prevalence. Thus, PPV and NPV can be tailored to particular populations by adapting Equations (6) and (7) by including the prevalence in the new population of interest:

$$\text{PPV}_{\text{new}_L} = \frac{\text{sensitivity}_{\text{MA}} \times \text{prevalence}_L}{[\text{sensitivity}_{\text{MA}} \times \text{prevalence}_L] + [(1 - \text{specificity}_{\text{MA}})(1 - \text{prevalence}_L)]} \quad (9)$$

$$\text{NPV}_{\text{new}_L} = \frac{\text{specificity}_{\text{MA}} \times (1 - \text{prevalence}_L)}{[(1 - \text{sensitivity}_{\text{MA}}) \times \text{prevalence}_L] + [\text{specificity}_{\text{MA}} \times (1 - \text{prevalence}_L)]} \quad (10)$$

where prevalence_L is the known prevalence in the intended population. If this prevalence is unknown, it might be substituted by the observed prevalence from a sample of patients from the intended population, to use as a proxy for the true prevalence.

4.1.3. Modelling option C: fit a bivariate meta-analysis of PPV and NPV, with prevalence as a covariate. Rather than deriving PPV and NPV by considering sensitivity and specificity independent of prevalence, a more sophisticated option is to consider sensitivity, specificity and prevalence jointly. Chu *et al.* [30] propose a trivariate meta-analysis for this purpose. However, in our experience, it is difficult to estimate reliably the two between-study correlations in this model. Thus, we take the approach suggested by Leeftang *et al.* [5], where PPV and NPV (rather than sensitivity and specificity) are now the two measures of interest, and a covariate included for prevalence. This bivariate meta-regression is applied to the available cohort studies in the meta-analysis and can be written as

$$\begin{aligned} r_{11i} &\sim \text{Binomial}(n_{+i}, \text{PPV}_i) \\ r_{00i} &\sim \text{Binomial}(n_{-i}, \text{NPV}_i) \\ \begin{pmatrix} \text{logit}(\text{PPV}_i) \\ \text{logit}(\text{NPV}_i) \end{pmatrix} &\sim N \left[\begin{pmatrix} \delta_1 + \gamma_1 \text{prevalence}_i \\ \delta_0 + \gamma_0 \text{prevalence}_i \end{pmatrix}, \mathbf{\Omega} \right], \mathbf{\Omega} = \begin{pmatrix} \tau_{\text{PPV}}^2 & \tau_{\text{PPV}} \tau_{\text{NPV}} \rho \\ \tau_{\text{PPV}} \tau_{\text{NPV}} \rho & \tau_{\text{NPV}}^2 \end{pmatrix} \end{aligned} \quad (11)$$

Here, n_{+i} and n_{-i} denote the number who tested positive and negative, respectively, in study i . The model specifies that the true $\text{logit}(\text{PPV}_i)$ and $\text{logit}(\text{NPV}_i)$ in each study depends on the prevalence (prevalence_i) of that study (note that an alternative specification could be to use $\text{logit}(\text{prevalence}_i)$ as the covariate, although for brevity, this is not considered further here). It allows for unexplained between-study heterogeneity in $\text{logit}(\text{PPV}_i)$ and $\text{logit}(\text{NPV}_i)$, τ_{PPV}^2 and τ_{NPV}^2 , respectively, and their potential between-study correlation (ρ). A disadvantage of model (11) is that it discards studies (e.g. case-control studies) that do not allow reliable estimation of PPV and NPV but could otherwise contribute towards sensitivity and specificity.

Model (11) can be fitted using the same techniques as described for model (1), and the estimated parameters enable PPV and NPV to be derived in a new population (L) if the prevalence (prevalence_L) in the intended population is specified, as follows:

$$\text{PPV}_{\text{new}_L} = \frac{\exp(\hat{\delta}_1 + \hat{\gamma}_1 \text{prevalence}_L)}{1 + \exp(\hat{\delta}_1 + \hat{\gamma}_1 \text{prevalence}_L)} \quad (12)$$

$$\text{NPV}_{\text{new}_L} = \frac{\exp(\hat{\delta}_0 + \hat{\gamma}_0 \text{prevalence}_L)}{1 + \exp(\hat{\delta}_0 + \hat{\gamma}_0 \text{prevalence}_L)} \quad (13)$$

4.2. Internal–external cross-validation for checking calibration of predicted PPV and NPV in new populations

Whatever strategy is used to derive PPV and NPV in new populations, external validation is required to check the calibration of predictions with observed risk. However, it is well known that calibration performance of predictive probabilities is optimistic when they are estimated and then tested on the same data [26, 31, 32]. Thus, to validate the chosen strategy, we suggest using internal–external cross-validation, which omits a study from the meta-analysis that derives the prediction equations, so that calibration can be checked in independent data. The process is then repeated, with a different study omitted in each cycle. The approach was proposed by Royston *et al.* [27] and recently extended by Debray *et al.* [28] for validation of multivariable prediction models. We now implement it within the context of test accuracy studies and extend it to include a meta-analysis that summarises the calibration performance across all cycles.

4.2.1. Summarising overall calibration performance. Suppose that the focus is on summarising the *overall* calibration between predicted and observed outcome risks in new populations. The internal–external cross-validation approach proceeds as follows:

- (1) Select a study L to be omitted
- (2) In the remaining $k - 1$ studies, fit a chosen meta-analysis model and use a chosen strategy (e.g. options A to C earlier) to produce equations for deriving PPV and NPV in a new population. Apply these equations to the omitted study L , to calculate the predicted PPV_{new_L} and NPV_{new_L} .
- (3) In the omitted study L , calculate the total observed (O) and the total expected (E) outcomes, where

$$O_L = n_{1L}$$

$$E_L = PPV_{new_L}(r_{11L} + (n_{0L} - r_{00L})) + (1 - NPV_{new_L})(r_{00L} + (n_{1L} - r_{11L})) \quad (14)$$

where PPV_{new_L} and NPV_{new_L} are those calculated using the equations derived in step (2).

- (4) In the omitted study, compute the *overall calibration* of predicted probabilities by $(O_L/E_L) = (O/E)_L$, and calculate $\ln(O/E)_L$ and its standard error, s_L .
Defining $P_{OL} = O_L / (n_{1L} + n_{0L})$ and $P_{EL} = E_L / (n_{1L} + n_{0L})$, we can write

$$\ln(O/E)_L = \ln(P_{OL}/P_{EL}) = \ln(P_{OL}) - \ln(P_{EL}),$$

with P_{OL} and P_{EL} the observed and expected (O/E) proportion with the outcome in the omitted study, respectively. Treating the expected proportion P_{EL} as fixed, the standard error of $\ln(O/E)_L$ equals the standard error of $\ln(P_{OL})$, which is given approximately by [33]

$$s_L = \sqrt{\frac{1 - P_{OL}}{(n_{1L} + n_{0L})P_{OL}}} \quad (15)$$

- (5) Repeat steps (1) to (4) for each omitted study, giving a set of k values for $\ln(O/E)_L$ and s_L .
- (6) Perform a (frequentist or Bayesian) random-effects meta-analysis of the $\ln(O/E)_L$ values as follows:

$$\ln(O/E)_L \sim N(\theta_L, s_L^2)$$

$$\theta_L \sim N(\beta, \tau^2) \quad (16)$$

- (7) Summarise calibration across all studies by displaying the meta-analysis results using a forest plot including the summary O/E value, its 95% CI, I^2 and a 95% prediction interval for the O/E value in a new population.

The omitted studies in step (1) must be cohort studies (or cross-sectional studies for diagnostic tests). Other studies, such as case-control studies, that do not provide unbiased estimates of outcome risk (or disease prevalence for diagnostic test studies) might contribute to the developed prediction equations in step (2) (e.g. using options A or B in Section 4.1).

Compared with previous use of the internal–external cross-validation approach [27, 28], we have added the meta-analysis component in steps (6) and (7). Because uncertainty in each ratio O/E reflects only within-study error for the omitted study and not the error associated with computing the expected values from the other studies, it is appropriate to combine these across studies using the within-study errors in the weights. This allows us to quantify whether observed differences in overall calibration are due to chance

or heterogeneity and to summarise calibration performance on average and in potential new populations. Perfect calibration would be revealed by a summary O/E value of 1 (indicating perfect calibration on average across all populations), together with no between-study heterogeneity in calibration performance, resulting in a narrow 95% prediction interval for the O/E in a new population. Such perfection is unlikely, and so researchers might consider what calibration performance would be acceptable. For example, a 95% prediction interval for O/E falling between 0.9 and 1.1 might be deemed acceptable, as the true risk is then within 10% of the predicted risk in new populations. In other words, one desires a large probability (0.95 or more) that O/E is between 0.9 and 1.1 in new population.

4.2.2. Summarising calibration of PPV and NPV separately and more exactly. The earlier analyses allow the *overall* calibration to be examined, comparing the total observed outcome events across both test-positive and test-negative individuals with the total expected number. Also of interest is the calibration of PPV and NPV themselves, especially as one might be more clinically important than the other. For example, in the thyroidectomy example, a reliable NPV is crucial as those with a negative PTH test may be discharged early to release valuable resources (such as beds and clinical care) to the hospital. The calibration of PPV and NPV can be undertaken using the same approach as described in Section 4.2.1 but with a separate O/E estimated for test-positive and test-negative patients in each study, followed by a separate meta-analysis of O/E for each.

However, when considering NPV (or PPV) separately, it is increasingly likely that some studies will have no events (or no non-events) for test-negative (or test-positive) patients. In such studies, the standard error of $\ln(\text{O/E})_L$ cannot be derived using Equation (15) as P_{OL} is 1 (or 0), and thus, meta-analysis model (16) is problematic. A more exact meta-analysis is thus proposed, which models the binomial likelihood directly in each study to avoid having to derive standard errors and thereby accommodate studies with zero events (or zero non-events) [13]. Without loss of generality, consider just PPV so that the analysis includes only test-positive ('+') patients. The model is written as

$$\begin{aligned} r_{11L} &\sim \text{Binomial}(n_{+L}, \text{PPV}_L) \\ \text{logit}(\text{PPV}_L) &= \alpha_L + \text{logit}(\text{PPVnew}_L) \\ \alpha_L &\sim N(a, \tau_a^2) \end{aligned} \quad (17)$$

where $\text{logit}(\text{PPVnew}_L)$ is an offset term corresponding to the logit of the predicted PPV for study L obtained from step (2). n_{+L} is the total number of test-positive patients in study L , and r_{11L} is the number of those who truly have the outcome. The α_L are often referred to as the calibration-in-the-large [26], and so the estimate of a provides the summary calibration-in-the-large. A more formal interpretation is that a is the average log odds ratio, summarising the log of observed to expected odds across studies. A value of 0 thus indicates perfect calibration. The heterogeneity, τ_a^2 , is the between-study variation in calibration, and this will be zero if calibration performance is consistent across all populations. Model (17) can be fitted in either a Bayesian or a frequentist framework (Supporting Information S5 and S6), with posterior (predictive) inferences then derived. For example, following frequentist estimation and given a particular PPVnew_L , an approximate 95% prediction interval for the true logit PPV is

$$\left(\hat{a} - t_{k-2,0.975} \sqrt{\hat{\tau}_a^2 + \text{Var}(\hat{a})} \right) + \text{logit}(\text{PPVnew}_L), \left(\hat{a} + t_{k-2,0.975} \sqrt{\hat{\tau}_a^2 + \text{Var}(\hat{a})} \right) + \text{logit}(\text{PPVnew}_L) \quad (18)$$

The lower and upper bounds can then be transformed back to the PPV scale, or the O/E scale, as desired.

4.3. Applications

We now apply the internal-external cross-validation approach to the two examples introduced in Section 2. The results are summarised in Tables IV–VII, and Figures 2–5.

4.3.1. Ear temperature for diagnosis of fever. In Section 3, the meta-analysis revealed that ear temperature above or below 38 °C only moderately discriminated those with and without fever. We now use the internal-external cross-validation approach to evaluate the calibration of post-test probabilities, PPV and NPV, derived using options A to C. The results at each cycle of the procedure are given in Table IV for each strategy, and Figure 2 and Tables V and VI summarise calibration performance.

Table IV. Results of the internal–external cross-validation procedure for the temperature data.

| Study | Observed no. with fever (O) | Summary prevalence from meta-analysis | | | | Option A Post-test predictions obtained using summary sensitivity, summary specificity and summary prevalence (Equations (6) and (7)) | | | | Option B Post-test predictions obtained using summary sensitivity, summary specificity and observed prevalence (Equations (9) and (10)) | | | | Option C Post-test predictions obtained after bivariate meta-regression of PPV and NPV on prevalence (Equations (12) and (13)) | | | |
|---------|-----------------------------------|---------------------------------------|------------------------|---------------------------------------|---------------------------------------|---|------------------|-------------------------|------|---|------------------|-------------------------|------|--|------------------|-------------------------|------|
| | | analysis model (8)* | analysis model (1)* | sensitivity analysis model (1)* | specificity analysis model (1)* | Expected | | | | Expected | | | | Expected | | | |
| | | Predicted PPV | Predicted NPV | no. with fever ^ (E) | O/E | Predicted PPV | Predicted NPV | no. with fever ^ (E) | O/E | Predicted PPV | Predicted NPV | no. with fever ^ (E) | O/E | Predicted PPV | Predicted NPV | no. with fever ^ (E) | O/E |
| Brennan | 203 | 0.55 | 0.45 | 0.644 | 0.99 | 0.97 | 0.77 | 207.59 | 0.98 | 0.98 | 0.69 | 226.11 | 0.90 | 0.98 | 0.75 | 211.83 | 0.96 |
| Davis | 18 | 0.27 | 0.48 | 0.661 | 0.98 | 0.97 | 0.76 | 24.41 | 0.74 | 0.94 | 0.89 | 16.85 | 1.07 | 0.96 | 0.78 | 22.71 | 0.79 |
| Green | 9 | 0.43 | 0.46 | 0.631 | 0.98 | 0.97 | 0.76 | 10.92 | 0.82 | 0.97 | 0.78 | 10.58 | 0.85 | 0.96 | 0.76 | 10.80 | 0.83 |
| Greenes | 109 | 0.36 | 0.46 | 0.67 | 0.98 | 0.97 | 0.77 | 110.91 | 0.98 | 0.96 | 0.83 | 94.12 | 1.16 | 0.95 | 0.78 | 106.11 | 1.03 |
| Hoffman | 42 | 0.42 | 0.47 | 0.645 | 0.99 | 0.97 | 0.77 | 47.23 | 0.89 | 0.96 | 0.80 | 44.56 | 0.94 | 0.97 | 0.76 | 47.06 | 0.89 |
| Hooker | 15 | 0.38 | 0.46 | 0.65 | 0.98 | 0.97 | 0.76 | 16.63 | 0.90 | 0.96 | 0.82 | 14.97 | 1.00 | 0.96 | 0.78 | 16.07 | 0.93 |
| Lanham | 103 | 0.58 | 0.47 | 0.666 | 0.98 | 0.97 | 0.76 | 81.98 | 1.26 | 0.98 | 0.67 | 94.01 | 1.10 | 0.98 | 0.76 | 82.71 | 1.25 |
| Muma | 87 | 0.39 | 0.46 | 0.662 | 0.98 | 0.97 | 0.78 | 84.73 | 1.03 | 0.96 | 0.83 | 75.98 | 1.15 | 0.95 | 0.78 | 84.66 | 1.03 |
| Nypaver | 425 | 0.48 | 0.45 | 0.652 | 0.98 | 0.97 | 0.78 | 408.91 | 1.04 | 0.97 | 0.76 | 426.06 | 1.00 | 0.97 | 0.76 | 421.87 | 1.01 |
| Rhoads | 27 | 0.42 | 0.47 | 0.683 | 0.98 | 0.96 | 0.77 | 20.25 | 1.33 | 0.96 | 0.80 | 18.12 | 1.49 | 0.96 | 0.77 | 19.78 | 1.36 |
| Stewart | 59 | 0.75 | 0.43 | 0.65 | 0.98 | 0.97 | 0.78 | 59.88 | 0.99 | 0.99 | 0.49 | 67.79 | 0.87 | 0.98 | 0.56 | 65.61 | 0.90 |

*Obtained from meta-analysing all studies excluding the study in the row.

^Expected number for the study in the row, based on using Equation (14) and the positive predictive value (PPV) and negative predictive value (NPV) in the prior two columns.

Table V. Meta-analysis results for overall calibration (O/E) when using model (16), as estimated in a frequentist or Bayesian framework.[†]

| Example | Predicted PPV and NPV obtained using ... | Statistical framework | Summary O/E (95% CI) | $\hat{\tau}$ (95% CI) | 95% prediction interval for O/E in a new population | Probability $0.9 < \text{O/E} < 1.1$ in a new population |
|------------------|--|-----------------------|----------------------|-----------------------|---|--|
| Temperature data | Option A | Bayesian | 1.02 (0.93 to 1.11) | 0.10 (0.01 to 0.23) | 0.78 to 1.31 | 0.67 |
| | Option A | Frequentist | 1.02 (0.95 to 1.10) | 0.08 | 0.85 to 1.24 | — |
| PTH data 1–2 h | Option B | Bayesian | 1.02 (0.73 to 1.38) | 0.14 (0.01 to 0.45) | 0.56 to 1.74 | 0.40 |
| | Option B | Frequentist | 1.01 (0.79 to 1.29) | 0 | 0.68 to 1.50 | — |

[†]All frequentist analyses used method of moments to estimate the model.

All Bayesian analyses used a prior $N(0, 1\ 000\ 000)$ for the mean $\ln(\text{O/E})$ and a prior uniform(0, 0.25) for τ , with a 10 000 burn-in followed by 100 000 samples for posterior inferences.

O/E, observed/expected; PPV, positive predictive value; NPV, negative predictive value.

| Table VI. Meta-analysis results for calibration of either PPV or NPV, when using meta-analysis model (17) as estimated in a frequentist or Bayesian framework. [†] | | | | | | |
|---|----------------------|-----------------------|-----------------------------------|---------------------|--|--|
| Example | Meta-analysis method | Statistical framework | Summary calibration, a (95% CI) | $\hat{\tau}_a$ | 95% prediction interval for O/E in a new population* | Probability $0.9 < O/E < 1.1$ in a new population* |
| Calibration for just NPV PTH data 1–2 h | Option B | Bayesian | 0.24 (–0.97 to 1.81) | 0.51 (0.03 to 0.98) | 0.86 to 1.05 | 0.95 |
| | Option B | Frequentist | 0.093 (–1.06 to 1.25) | 0 | 0.87 to 1.03 | — |
| | Option B | Bayesian | 0.021 (–0.82 to 1.02) | 0.51 (0.03 to 0.97) | 0.86 to 1.04 | 0.95 |
| | Option B | Frequentist | –0.044 (–0.83 to 0.74) | 0.34 | 0.80 to 1.03 | — |
| Calibration for just PPV Temperature data | Option A | Bayesian | –0.017 (–0.63 to 0.77) | 0.65 (0.11 to 0.99) | 0.90 to 1.03 | 0.98 |
| | Option A | Frequentist | 0.015 (–0.75 to 0.78) | 0.70 | 0.87 to 1.03 | — |

[†]All frequentist analyses used maximum likelihood estimation of model (17). All Bayesian analyses used a prior distribution of $N(0, 1\ 000\ 000)$ for a , and a prior distribution of $\text{uniform}(0, 1)$ for τ , with a 10 000 burn-in followed by 100 000 samples for posterior inferences. Median values of the posterior distribution are shown for a and τ .

*Based on a predicted positive predictive value (PPV) of 0.97 in the temperature analysis, and a negative predictive value (NPV) of 0.95 in the parathyroid (PTH) analysis.

O/E, observed/expected.

Table VII. Results of the internal-external cross-validation procedure for the PTH data.

| Study | Observed no. with fever (O) | Summary | | | | | Option A Post-test predictions obtained using summary sensitivity, summary specificity and summary prevalence (Equations (6) and (7)) | | | | | Option B Post-test predictions obtained using summary sensitivity, summary specificity and observed prevalence (Equations (9) and (10)) | | | | |
|------------------------------|-----------------------------------|----------------------------------|------------------------|--------------------------|------------|--|---|-------|--------------------------------------|----------|------|---|--------------------------------------|-----|--|--|
| | | prevalence from meta-analysis | | Summary meta-analysis | | Summary meta-analysis model (1)* | Expected | | | Expected | | | | | | |
| | | model (8)* | Observed prevalence | model (1)* | model (1)* | | PPV | NPV | Predicted no. with outcome^(E) | O/E | PPV | NPV | Predicted no. with outcome^(E) | O/E | | |
| | | | | | | | | | | | | | | | | |
| (1) PTH measured at 1–2 h | | | | | | | | | | | | | | | | |
| Lam | 12 | 0.32 | 0.26 | 0.91 | 0.78 | 0.60 | 0.96 | 9.37 | 1.28 | 0.66 | 0.95 | 10.51 | 1.14 | | | |
| Lombardi | 16 | 0.31 | 0.26 | 0.93 | 0.80 | 0.62 | 0.97 | 14.01 | 1.14 | 0.68 | 0.96 | 15.64 | 1.02 | | | |
| McLeod | 12 | 0.32 | 0.26 | 0.94 | 0.82 | 0.65 | 0.97 | 11.77 | 1.02 | 0.72 | 0.97 | 13.11 | 0.92 | | | |
| Warren, 2002 | 2 | 0.20 | 0.28 | 0.95 | 0.82 | 0.67 | 0.98 | 2.21 | 0.90 | 0.57 | 0.99 | 1.82 | 1.10 | | | |
| Warren, 2004 | 3 | 0.12 | 0.31 | 0.93 | 0.83 | 0.71 | 0.96 | 7.22 | 0.42 | 0.42 | 0.99 | 3.99 | 0.75 | | | |
| (2) PTH measured at 0–20 min | | | | | | | | | | | | | | | | |
| Lo | 11 | 0.11 | 0.24 | 0.75 | 0.83 | 0.59 | 0.91 | 33.60 | 0.33 | 0.36 | 0.96 | 18.82 | 0.58 | | | |
| Lombardi | 16 | 0.31 | 0.16 | 0.81 | 0.76 | 0.39 | 0.96 | 8.32 | 1.92 | 0.60 | 0.90 | 14.12 | 1.13 | | | |
| McLeod | 13 | 0.23 | 0.18 | 0.87 | 0.81 | 0.50 | 0.97 | 11.15 | 1.17 | 0.58 | 0.95 | 13.22 | 0.98 | | | |
| Warren, 2002 | 4 | 0.25 | 0.18 | 0.83 | 0.78 | 0.47 | 0.95 | 2.94 | 1.36 | 0.56 | 0.93 | 3.69 | 1.08 | | | |
| Warren, 2004 | 3 | 0.12 | 0.21 | 0.84 | 0.77 | 0.49 | 0.95 | 3.70 | 0.81 | 0.32 | 0.97 | 2.26 | 1.33 | | | |

*Obtained from meta-analysing all studies excluding the study in the row.

[^]Expected number for the study in the row, based on using Equation (14) and the positive predictive value (PPV) and negative predictive value (NPV) in the prior two columns. PTH, parathyroid.

Overall calibration. First consider the overall calibration of observed to expected values, across all patients, for the case where PPV and NPV are derived by option A (use the summary sensitivity, specificity and prevalence from meta-analysis of the other studies) (Table IV). The predicted PPV and NPV are close to 0.97 and 0.77 for most studies (Table IV). Frequentist estimation of meta-analysis model (16), using methods of moments, gives a summary O/E close to 1 with a narrow CI (summary O/E = 1.02, 95% CI: 0.95 to 1.10), suggesting excellent calibration on average in new populations (Figure 2(a); Table V). However, there is heterogeneity in calibration performance ($I^2 = 51\%$), a likely consequence of the heterogeneity in sensitivity and specificity identified in Section 3. A 95% prediction interval for the O/E in a new population is 0.85 to 1.24, suggesting that the true fever risk will be between 24% higher and 15% lower than that expected in new populations. Bayesian estimation gives similar inferences, although with wider intervals, and there is a probability of 0.66 that $0.9 < \text{O/E} < 1.1$ (Table V), and so in two-thirds of populations, calibration will reassuringly be within 10% of that predicted.

Interestingly, calibration appears worse when the internal–external validation approach is repeated with PPV and NPV calculated using option B (prevalence taken from the omitted study's population, combined with summary sensitivity and specificity). As true population prevalences were unknown in omitted studies, we mimicked this situation by assuming the observed prevalence in the omitted study was actually the true prevalence. The forest plot summarising O/E for this approach is shown in Figure 2(b). Compared with the previous analysis, heterogeneity in calibration performance increases ($I^2 = 62\%$), and thus, the 95% prediction interval is wider (0.81 to 1.30). Therefore, post-test predictions appear less reliable using option B rather than option A. Option C (bivariate meta-regression of PPV and NPV on prevalence) performs similarly to option A. The summary O/E is slightly improved (1.01), but there is additional heterogeneity ($I^2 = 56\%$), and thus, the 95% prediction interval is similar (0.82 to 1.25).

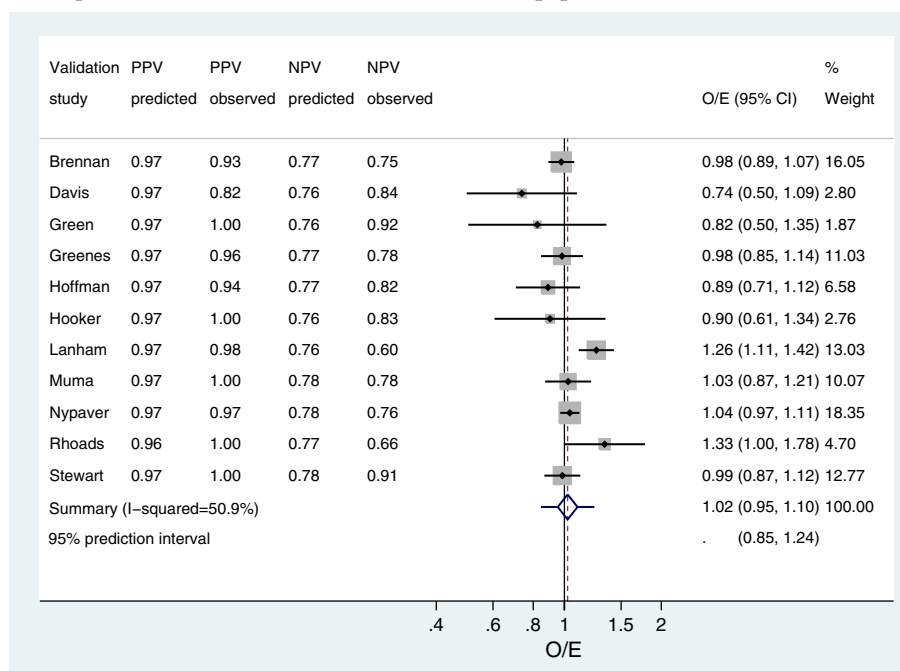
Calibration of PPV. Because of NPV being too low to rule out fever, one might examine just the calibration of PPV. Using option A to derive PPV in new populations, the predicted PPV is close to 0.97 in all cycles of the internal–external cross-validation approach (Table IV). Some studies did not have any test-positive children without fever, and thus, the exact meta-analysis model (17) was used. Maximum likelihood estimation gives a summary calibration, a , of 0.015 (95% CI: -0.75 to 0.78) (Table VI). This reveals that, on average, there is close to perfect agreement between expected and observed PPVs in the studies. However, there is heterogeneity ($\tau_a = 0.70$) in calibration performance, which, when using Equation (18) followed by back transformations, corresponds to a 95% prediction interval of 0.87 to 1.03 for the O/E in a new population where the predicted PPV was 0.97. Bayesian estimates are very similar (Table VI), and there is a large probability of 0.98 that O/E will be between 0.9 and 1.1 in a new population. Further, for a predicted PPV of 0.97, there is a probability of 0.78 that the true PPV will be between 0.95 and 1. The calibration between observed and predicted PPVs using option A is shown in Figure 4(a). Calibration performance is not improved when deriving PPV using options B or C (results not shown), as either the summary calibration performance is worse or the heterogeneity is larger.

Thus, PPV derived from option A appears to calibrate extremely well in new populations. We thus derived a final PPV from option A to be used in clinical practice, obtained by placing into Equation (6) the summary estimates of sensitivity, specificity and prevalence from the meta-analyses of all 11 studies. This gave a high PPV of 0.97, and thus, an ear temperature $\geq 38.0^\circ\text{C}$ strongly indicates that a child truly has a fever.

4.3.2. PTH for predicting hypocalcaemia. Section 3.3 identified that PTH has high sensitivity and specificity at a threshold of 65% when PTH is measured 1–2 h post-surgery (summary sensitivity, 0.93 and summary specificity, 0.81). For this test, we now use the internal–external cross-validation approach to examine and compare the calibration of PPV and NPV in new populations as derived from options A and B. Given there were only five studies in the meta-analysis, the meta-regression approach of option C was not deemed sensible and is thus not evaluated here.

Overall calibration. NPV and PPV are both estimated consistently high during the internal–external cross-validation process (Table VII). When post-test probabilities are based on option A (using the summary sensitivity, specificity and prevalence), frequentist estimation of model (16) indicates there is no heterogeneity in calibration performance ($I^2 = 0\%$) (Figure 3(a)). The summary O/E is 1.07, suggesting under-prediction of hypocalcaemia events in new populations. When the post-test probabilities are

(a) Option A used to derive PPV and NPV in new populations



(b) Option B used to derive PPV and NPV in new populations

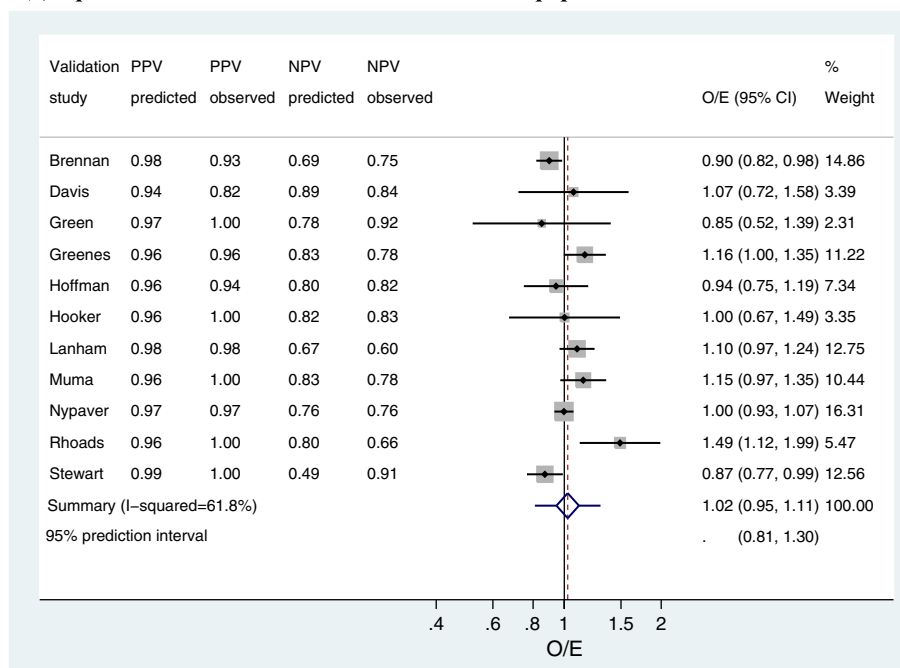


Figure 2. Meta-analysis of the observed/expected (O/E) calibration statistics (frequentist estimation of model (16)) from the internal-external cross-validation approach applied to the ear temperature data for diagnosis of fever. PPV, positive predictive value; NPV, negative predictive value.

derived using option B (summary sensitivity and specificity, combined with observed prevalence in omitted study), there continues to be no heterogeneity, but the summary O/E is improved to 1.01, indicating almost perfect calibration on average (Figure 3(b)). Although there is no estimated heterogeneity, the prediction interval (0.68 to 1.50) is wide (Figure 3(b)), as it accounts for large uncertainty in the heterogeneity and pooled estimates, a consequence of only five studies in the meta-analysis. Bayesian posterior

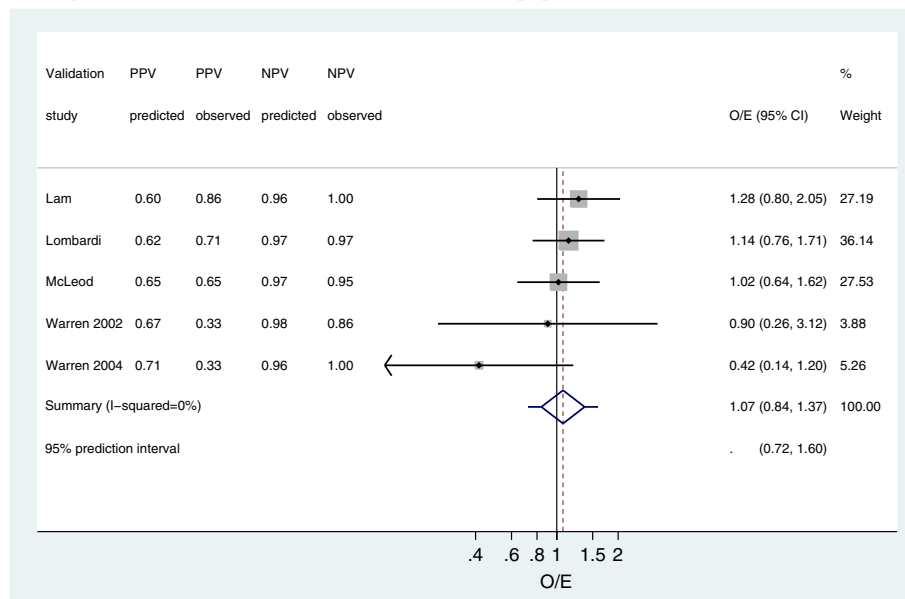
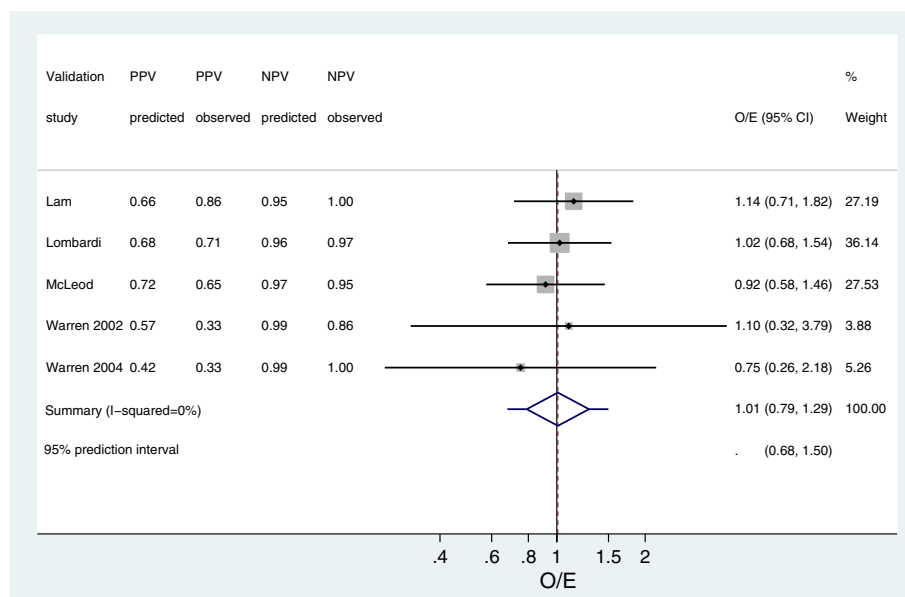
(a) Option A used to derive PPV and NPV in new populations*(b) Option B used to derive PPV and NPV in new populations*

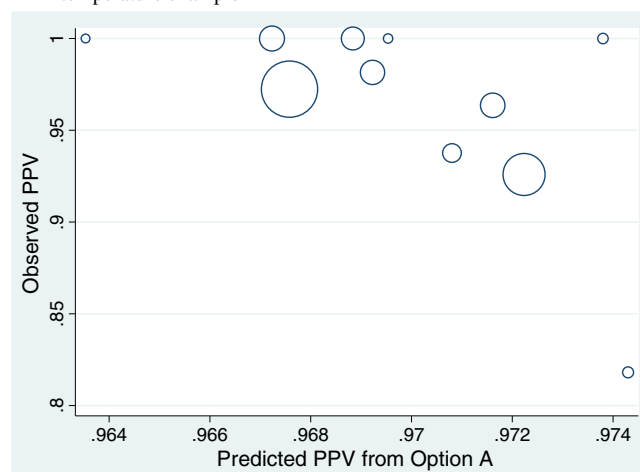
Figure 3. Meta-analysis of the observed/expected (O/E) calibration statistics (frequentist estimation of model (16)) from the internal–external cross-validation approach applied to the parathyroid data at 1–2 h for prediction of hypocalcaemia. PPV, positive predictive value; NPV, negative predictive value.

estimates are very similar, although intervals are even wider. There is a probability of only 40% that the true O/E is between 0.9 and 1.1, again reflecting the large uncertainty in the meta-analysis.

Calibration of NPV. As mentioned, the pivotal element for the PTH test is good calibration for NPV so that patients are not wrongly discharged from hospital. Table VII shows that the predicted NPV is consistently high in the cycles of internal–external cross-validation, ranging from 0.96 to 0.98 when the summary prevalence is used (option A) and from 0.95 to 0.99 when the observed population prevalence is used (option B). To examine calibration performance of predicted NPV in new populations, meta-analysis model (17) was applied to accommodate studies that had no test-negative patients with hypocalcaemia.

For predicted NPV derived using option B, the summary calibration, α , is 0.093 following frequentist estimation, which suggests only a small positive miscalibration between observed and predicted NPVs.

(a) Calibration of observed and predicted PPV values for the temperature example



(b) Calibration of observed and predicted NPV values for the PTH test measured 1-2 hours post-surgery

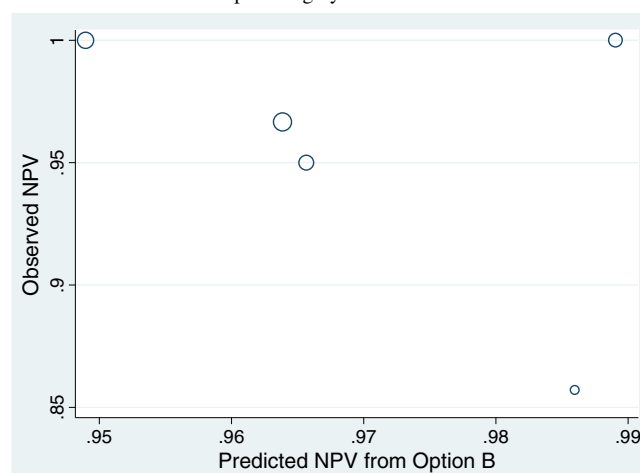


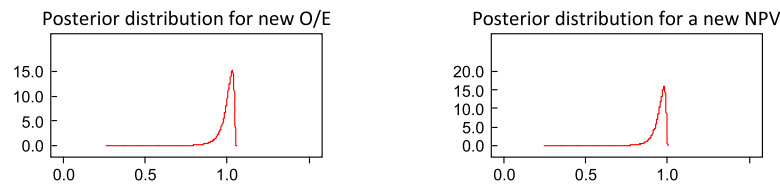
Figure 4. Calibration of predicted and observed post-test probabilities, for (a) positive predictive value (PPV) derived using option A in the temperature example and (b) negative predictive value (NPV) derived using option B in the parathyroid example. Each circle represents a study and is proportional to the study sample size.

There is also no estimated heterogeneity in calibration performance. For a predicted NPV of 0.95, these results correspond to a 95% prediction interval for the true NPV in a new population of 0.78 to 0.99, or an O/E of 0.87 to 1.03. These intervals are wide because of the small number of studies. For the same reason, Bayesian estimation is especially sensitive to the choice of prior distribution for τ , and frequentist and Bayesian results are less similar here than for the temperature analyses (Table VI). However, the Bayesian approach agrees that option B gives well-calibrated NPVs in new populations. For example, when using a $U(0,1)$ prior for τ , there is a large probability of 0.95 that option B will give a predicted NPV that calibrates within 10% of the true NPV in a new population (Table VI). The calibration of observed and predicted NPVs is shown in Figure 4(b). Predictive (posterior) distributions for the NPV and O/E in a new population are shown in Figure 5. Option A performs very similarly to option B.

Thus, although further studies would be useful to reduce the width of prediction intervals, the current evidence strongly suggests that option B (or even A) will give predicted NPVs that calibrate well in new populations. Using option B and deriving summary sensitivity and specificity from a meta-analysis of all 5 studies, the final prediction equation for a new NPV is

$$\text{NPV}_{\text{new}_L} = \frac{0.81 \times (1 - \text{prevalence}_L)}{[(1 - 0.93) \times \text{prevalence}_L] + [0.81 \times (1 - \text{prevalence}_L)]}$$

(a) Posterior (predictive) distributions for the performance of the PTH test at 1–2 hours to diagnose hypocalcaemia, assuming the predicted NPV is 0.95 as obtained from option B



(b) Posterior (predictive) distributions for the performance of the ear temperature test to diagnose fever, assuming the predicted PPV is 0.97 as obtained from option A

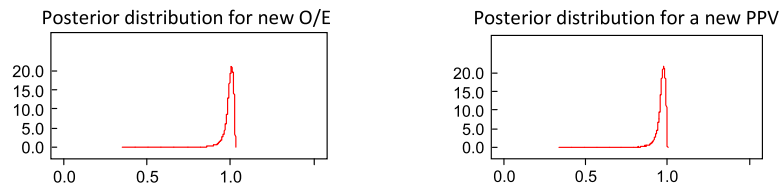


Figure 5. Posterior distributions for the true observed/expected (O/E), positive predictive value (PPV) or negative predictive value (NPV) in a new population, derived from Bayesian estimation of model (17).

5. Extensions

We now briefly consider two potentially useful extensions of our work.

5.1. Comparing test performance

The examination of calibration and discrimination performance in new populations is also a novel way to compare the performance of two different tests, or for comparing the same test at different thresholds or measurement methods. For example, in the PTH example, there are two competing tests: PTH measured at 0–20 min or at 1–2 h. Both have a high discrimination indicated by c statistics above 0.80 (see, e.g. Supporting Information S4), but which will perform better in new populations? For illustration, we now evaluate this for a threshold of 65%.

5.1.1. Comparison of sensitivity and specificity at a 65% threshold. For PTH at 0–20 min, fitting model (1) in a Bayesian framework gives a summary sensitivity of 0.82 (95% CI: 0.64 to 0.94) and a summary specificity of 0.79 (0.67 to 0.89). The probability that sensitivity will be greater than 0.8 in a new population is 0.60, and for specificity, it is 0.44. The probability that both will be above 0.8 in a new population is just 0.20. In contrast, recall that for PTH measured at 1–2 h, there was a probability of 0.57 that both would be above 0.8 in a new population, because of the larger summary sensitivity (0.93) and specificity (0.81). Thus, the discrimination of the test is substantially better when measured at 1–2 h.

5.1.2. Comparison of the calibration of post-test probabilities at a 65% threshold. At 0–20 min post-surgery, the internal–external cross-validation approach suggests that NPV is consistently predicted to be above 0.90, whereas PPV is much lower (Table VII). Higher NPV and PPVs are generally predicted at 1–2 h (Table VII), because of the larger sensitivity and specificity, and thus are clearly preferred if they calibrate well in new populations.

When using option B to derive post-test predictions, at 0–20 min the summary O/E from meta-analysis model (16) is 0.96, which suggests the test slightly over-predicts hypocalcaemia events, and the 95% prediction interval is wide (0.61 to 1.50). In contrast, at 1–2 h post-surgery, the summary O/E was better at 1.01, and the 95% prediction interval was slightly narrower (0.68 to 1.50), although still wide.

Thus, in terms of both discrimination and calibration performances in new populations, the PTH test measured at 1–2 h appears preferable based on the evidence available.

5.2. Predictions involving additional study-level and patient-level covariates

We suggested options A to C as possible strategies to derive post-test predictions in new populations, based on the existing meta-analysis results. If internal–external cross-validation indicates these are all unsuitable, for example, because of poor calibration of predictions in new populations, then further strategies are needed to tailor predictions more accurately. For example, additional study-level covariates could be included alongside prevalence in bivariate meta-regression model (11), to reduce heterogeneity further. Or alternatively, study-level covariates (including prevalence) could be included in a bivariate meta-regression extension of model (1), to explain heterogeneity in sensitivity and specificity, and thereby allow sensitivity and specificity to be predicted in a new population conditional on particular covariate values. For instance, accounting for the country of the study or the timing of the test may be included. If individual participant data (IPD) are available, one could also develop and validate a multivariable prediction model that contains the test value as a patient-level covariate, alongside multiple patient-level and study-level variables. The use of meta-analysis to examine the performance of multivariable prediction models is increasingly of interest [34, 35].

6. Discussion

Where heterogeneity exists, summary meta-analysis results might not be fit for purpose as test accuracy in particular populations may differ considerably from the average performance. Meta-analysts therefore need to be clearer about if and how their test accuracy results can be implemented in clinical practice. In this article, we have proposed approaches to improve the reporting and translation of meta-analysis results for use in clinical practice. We have brought together a number of existing concepts. In particular, we suggest that meta-analysts should derive and report prediction intervals for the *c* statistic, sensitivity, specificity and O/E in new populations. Probability statements should also be made about the likelihood of acceptable test performance in new populations. Further, by omitting one study during each cycle of estimation, we suggest using internal–external cross-validation to allow calibration of post-test probabilities (PPV and NPV) to be examined in independent data on multiple occasions. This enables different strategies for calculating PPV and NPV to be formally evaluated and compared, to identify the best approach for deriving them in clinical practice.

Of course, the true impact of a diagnostic or prognostic test is ultimately measured on how it improves patient outcomes [36]. This is perhaps best evaluated through a randomised trial, where one group has access to test results and the other does not. Our framework can help prioritise which tests are worth investment of a trial, by revealing which tests perform consistently well in new populations in terms of discrimination and calibration of post-test probabilities.

6.1. Limitations of our work

The use of prediction intervals to summarise calibration and discrimination performance will be more informative when most studies have a reasonable sample size, and there are a reasonable number of studies available. When there are fewer than about 10 studies, predictive distributions and regions might be wide, as seen in the overall calibration results for the PTH test (Table V). With small numbers of studies, the between-study variance is estimated with large uncertainty, and this will be propagated in subsequent prediction intervals and distributions. Frequentist and Bayesian inferences may also be less similar with few studies (see, e.g. PTH results in Table VI), as prior distributions become more informative.

Even with large numbers of studies, wide prediction intervals will arise when there is large between-study heterogeneity in test performance. This indicates the test may be useful in some populations but not in others, and so identifying those populations in which it works well becomes important (Section 5.2). However, identifying the causes of heterogeneity is also problematic, as meta-regression often has low power and is prone to ecological bias [37, 38].

Underpinning all our methods is the assumption that studies included in the meta-analysis are from relevant populations for which the test is desired and that the reference standard for true disease (or outcome) status is suitable in all studies. The broader the inclusion criteria (e.g. different patient pathways, different countries and different reference standards), the more likely heterogeneity in test performance will be identified. Between-study heterogeneity may also arise because of different methods of test measurement, implicit or explicit differences in the choice of threshold and the quality (risk of bias) of the studies [39]. This is clearly unwanted. Meta-analysis results are more applicable when studies are of good quality and are consistent in how the test is implemented. Indeed, prediction intervals are really

only relevant when based on studies with low risk of bias, as otherwise, they are widened to account for additional heterogeneity due to bias. Meta-analysts might therefore use a strict study inclusion criteria before applying the methods suggested in this paper. For brevity, we have not examined risk of bias for the examples in this paper, but future work may revisit this for these data.

A key proposal is to use internal–external cross-validation to examine the calibration of predicted PPV and NPV in new populations. For options B and C, these predicted values are derived in the external validation study with use of the known (expected) prevalence in that population. However, when implementing options B and C in both our applied examples, we did not know the known (expected) prevalence, and so used the observed prevalences. As we replaced the known prevalence with the observed prevalence, one could argue that the findings for options B and C in our applied examples may still be optimistic of true performance. However, given that the observed prevalence in a study should (for cohort studies) be an unbiased estimate of the true prevalence, we think this is perhaps a minor issue, although further research of this issue is appealing.

6.2. Related and further work

We believe that, if they are proposed for use in clinical practice, meta-analysis results about PPV and NPV (or meta-analysis equations to derive PPV and NPV in new populations) should be externally validated in new data. In other words, external validation is needed to *independently* check the reliability of post-test probabilities in target populations. This is currently not well discussed in the test accuracy meta-analysis literature. In contrast, in the prediction model field, the need for external validation is more widely recognised, as calibration and discrimination of multivariable model predictions are prone to over-optimism and may vary across populations. Indeed, in prediction model research, others have already considered meta-analysis of discrimination and calibration statistics from multiple external validation studies [24, 34, 35].

For test research, given data from multiple studies, we therefore proposed the internal–external cross-validation approach to evaluate calibration in external data, to avoid over-optimism [26]. This allows previously recommended strategies to derive PPV and NPV (e.g. based on meta-analysis models proposed by Leeflang *et al.* [5] and Chu *et al.* [30]) to be formally evaluated in independent data from new populations. We note that if one study is much larger than other studies, it is perhaps wise to always retain it for the meta-analysis (and thus never exclude it for external validation). In this manner, each cycle of internal–external cross-validation will retain the majority of the available data for obtaining the meta-analysis equation to predict PPV and NPV, and so the equation is likely to be very similar in each cycle. In related work, Willis and Hyde [10, 40] also suggest that test accuracy meta-analysis results should be tailored to new populations. They consider a probabilistic approach to study selection using information from the setting of interest; as such, only studies which are plausible for the setting are considered for meta-analysis. The performance of this approach could also be evaluated using the internal–external cross-validation framework.

An interesting issue is how to define ‘good’ discrimination and calibration performance. In our application, we considered the probability that a test’s calibration (O/E) is between 0.9 and 1.1, and the probability that a test’s discrimination c statistic is above 0.7 or 0.8 in new populations. Such criteria are subjective, but probability statements such as these allow multiple tests to be compared and thereby help identify the best test for use. Such test comparisons are best considered in the subset of studies that evaluate all the tests of interest, to ensure direct comparisons in the same sets of patients and populations [41].

In small samples, the normal sampling distribution for c_i in model (4) may be a poor approximation, especially if c_i estimates are close to 1. Other transformations may thus be considered in further work, such as $\text{logit}(c_i)$, although van Klaveren *et al.* [24] conclude that the original c statistic scale is preferred. Also, we note that the prediction intervals assume the true effects follow a (bivariate) normal distribution across studies. The validity of this assumption requires empirical evaluation, and other more flexible distributions may be required [42]. Finally, in our PTH example, the binary outcome was hypocalcaemia by 48 h and was known for every patient. For other prognostic test situations, the outcome may be longer-term and some patients may be censored before the time of interest. Thus, extension of our work to time-to-event data is required.

7. Conclusion

In conclusion, we have suggested approaches to improve the translation of test accuracy meta-analysis results for clinical practice. Prediction intervals and probability statements are important ways to summarise a test's discrimination and calibration performance in new populations. Further, internal-external cross-validation allows meta-analysts to identify a reliable strategy for how clinicians should derive post-test probabilities in their population.

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References

- Deeks JJ. Systematic reviews in health care: systematic reviews of evaluations of diagnostic and screening tests. *BMJ* 2001; **323**:157–162.
- Reitsma JB, Glas AS, Rutjes AW, Scholten RJ, Bossuyt PM, Zwinderman AH. Bivariate analysis of sensitivity and specificity produces informative summary measures in diagnostic reviews. *Journal of Clinical Epidemiology* 2005; **58**: 982–990.
- Harbord RM, Deeks JJ, Egger M, Whiting P, Sterne JA. A unification of models for meta-analysis of diagnostic accuracy studies. *Biostatistics* 2007; **8**:239–251.
- Chu H, Cole SR. Bivariate meta-analysis of sensitivity and specificity with sparse data: a generalized linear mixed model approach. *Journal of Clinical Epidemiology* 2006; **59**:1331–1332. author reply 1332–1333.
- Leeftang MM, Deeks JJ, Rutjes AW, Reitsma JB, Bossuyt PM. Bivariate meta-analysis of predictive values of diagnostic tests can be an alternative to bivariate meta-analysis of sensitivity and specificity. *Journal of Clinical Epidemiology* 2012; **65**:1088–1097.
- Harbord RM, Whiting P, Sterne JA, Egger M, Deeks JJ, Shang A, Bachmann LM. An empirical comparison of methods for meta-analysis of diagnostic accuracy showed hierarchical models are necessary. *Journal of Clinical Epidemiology* 2008; **61**:1095–1103.
- Noordzij JP, Lee SL, Bernet VJ, Payne RJ, Cohen SM, McLeod IK, Hier MP, Black MJ, Kerr PD, Richards ML, Lo CY, Raffaelli M, Bellantone R, Lombardi CP, Cohen JJ, Dietrich MS. Early prediction of hypocalcemia after thyroidectomy using parathyroid hormone: an analysis of pooled individual patient data from nine observational studies. *Journal of the American College of Surgeons* 2007; **205**:748–754.
- Higgins JP, Thompson SG, Spiegelhalter DJ. A re-evaluation of random-effects meta-analysis. *Journal of the Royal Statistical Society, Series A* 2009; **172**:137–159.
- Riley RD, Higgins JP, Deeks JJ. Interpretation of random effects meta-analyses. *BMJ* 2011; **342**:d549.
- Willis BH, Hyde CJ. Estimating a test's accuracy using tailored meta-analysis – how setting-specific data may aid study selection. *Journal of Clinical Epidemiology* 2014; **67**:538–546.
- Craig JV, Lancaster GA, Taylor S, Williamson PR, Smyth RL. Infrared ear thermometry compared with rectal thermometry in children: a systematic review. *Lancet* 2002; **360**:603–609.
- Dodd SR, Lancaster GA, Craig JV, Smyth RL, Williamson PR. In a systematic review, infrared ear thermometry for fever diagnosis in children finds poor sensitivity. *Journal of Clinical Epidemiology* 2006; **59**:354–357.
- Hamza TH, van Houwelingen HC, Stijnen T. The binomial distribution of meta-analysis was preferred to model within-study variability. *Journal of Clinical Epidemiology* 2008; **61**:41–51.
- Zeger SL, Liang KY, Albert PS. Models for longitudinal data: a generalized estimating equation approach. *Biometrics* 1988; **44**:1049–1060.
- Pinheiro JC, Bates DM. Approximations to the log-likelihood function in the nonlinear mixed-effects model. *Journal of Computational and Graphical Statistics* 1995; **4**:12–35.
- Statistical Software. Release 13.0. Stata Corporation. College Station: TX, 2013.
- Lunn DJ, Thomas A, Best N, Spiegelhalter D. WinBUGS – a Bayesian modelling framework: concepts, structure, and extensibility. *Statistics and Computing* 2000; **10**:325–337.
- Lambert PC, Sutton AJ, Burton PR, Abrams KR, Jones DR. How vague is vague? A simulation study of the impact of the use of vague prior distributions in MCMC. *Statistics in Medicine* 2005; **24**:2401–2428.
- Browne WJ, Draper D. Implementation and performance issues in the Bayesian and likelihood fitting of multilevel models. *Computational Statistics* 2000; **15**:391–420.
- Zwinderman AH, Bossuyt PM. We should not pool diagnostic likelihood ratios in systematic reviews. *Statistics in Medicine* 2008; **27**:687–697.
- Wei Y, Higgins JP. Bayesian multivariate meta-analysis with multiple outcomes. *Statistics in Medicine* 2013; **32**: 2911–2934.
- Bujkiewicz S, Thompson JR, Sutton AJ, Cooper NJ, Harrison MJ, Symmons DP, Abrams KR. Multivariate meta-analysis of mixed outcomes: a Bayesian approach. *Statistics in Medicine* 2013; **32**:3926–3943.

23. Harrell FE. *Regression Modeling Strategies, with Applications to Linear models, Logistic Regression, and Survival Analysis*. Springer: New York, 2001.
24. van Klaveren D, Steyerberg EW, Perel P, Vergouwe Y. Assessing discriminative ability of risk models in clustered data. *BMC Med Res Methodol* 2014; **14**:5.
25. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials* 1986; **7**:177–188.
26. Steyerberg EW. *Clinical Prediction Models: A Practical Approach to Development, Validation, and Updating*. Springer: New York, 2009.
27. Royston P, Parmar MKB, Sylvester R. Construction and validation of a prognostic model across several studies, with an application in superficial bladder cancer. *Statistics in Medicine* 2004; **23**:907–926.
28. Debray TP, Moons KG, Ahmed I, Koffijberg H, Riley RD. A framework for developing, implementing, and evaluating clinical prediction models in an individual participant data meta-analysis. *Statistics in Medicine* 2013; **32**:3158–3158.
29. Hemingway H, Croft P, Perel P, Hayden JA, Abrams K, Timmis A, Briggs A, Udumyan R, Moons KG, Steyerberg EW, Roberts I, Schroter S, Altman DG, Riley RD. Prognosis research strategy (PROGRESS) 1: a framework for researching clinical outcomes. *BMJ* 2013; **346**:e5595.
30. Chu H, Nie L, Cole SR, Poole C. Meta-analysis of diagnostic accuracy studies accounting for disease prevalence: alternative parameterizations and model selection. *Statistics in Medicine* 2009; **28**:2384–2399.
31. Steyerberg EW, Moons KG, van der Windt DA, Hayden JA, Perel P, Schroter S, Riley RD, Hemingway H, Altman DG. Prognosis research strategy (PROGRESS) 3: prognostic model research. *PLoS Med* 2013; **10**:e1001381.
32. Bleeker SE, Moll HA, Steyerberg EW, Donders ART, Derksen-Lubsen G, Grobbee DE, Moons KGM. External validation is necessary in, prediction research: a clinical example. *Journal of Clinical Epidemiology* 2003; **56**:826–832.
33. Selvin S. *Survival Analysis for Epidemiologic and Medical Research: A Practical Guide*. Cambridge University Press: New York, 2008.
34. Pennells L, Kaptoge S, White IR, Thompson SG, Wood AM. Assessing risk prediction models using individual participant data from multiple studies. *American Journal of Epidemiology* 2014; **179**:621–632.
35. Snell KIE, Hua H, Debray TP, Ensor J, Look MP, Moons KG, Riley RD. Multivariate meta-analysis of individual participant data helps externally validate the performance and implementation of a prediction model. *Journal of Clinical Epidemiology* 2015. (submitted).
36. Ferrante di Ruffano L, Hyde CJ, McCaffery KJ, Bossuyt PM, Deeks JJ. Assessing the value of diagnostic tests: a framework for designing and evaluating trials. *BMJ* 2012; **344**:e686.
37. Schmid CH, Stark PC, Berlin JA, Landais P, Lau J. Meta-regression detected associations between heterogeneous treatment effects and study-level, but not patient-level, factors. *Journal of Clinical Epidemiology* 2004; **57**:683–697.
38. Berlin JA, Santanna J, Schmid CH, Szczech LA, Feldman HI. Individual patient- versus group-level data meta-regressions for the investigation of treatment effect modifiers: ecological bias rears its ugly head. *Statistics in Medicine* 2002; **21**: 371–387.
39. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, Leeflang MM, Sterne JA, Bossuyt PM. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Annals of Internal Medicine* 2011; **155**: 529–536.
40. Willis BH, Hyde CJ. What is the test's accuracy in my practice population? Tailored meta-analysis provides a plausible estimate. *Journal of Clinical Epidemiology* 2014. DOI: 10.1016/j.jclinepi.2014.10.002.
41. Takwoingi Y, Leeflang MM, Deeks JJ. Empirical evidence of the importance of comparative studies of diagnostic test accuracy. *Annals of Internal Medicine* 2013; **158**:544–554.
42. Lee KJ, Thompson SG. Flexible parametric models for random-effects distributions. *Statistics in Medicine* 2008; **27**: 418–434.

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